

Testing the AFLP protocol to determine how genetic diversity may influence the climate change response of Pale Poppy (*Papaver alboroseum*) at Portage Glacier, Alaska

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INTRODUCTION

Pale poppy (*Papaver alboroseum*)

- White/pale pink flower
- Self-fertilization
- Native to Kamchatka, Alaska, the Yukon Territory of Canada, and British Columbia, Canada (See Figure 1)
- Found in areas affected by volcanic eruption and deglaciation
- 8 samples of pale poppy used for this study
- Specific area of study was in Site B of the Portage Glacier Valley in Alaska (Collet, 2005)

RESULTS

Size Standard	LIZ500		
Concentration	6.25%	12.5%	25%
Primer Set Colors (success rate):	1:8	7:8	6:8
Blue ECOR1-1 x MSE1-5			
Green ECOR1-2 x MSE1-2	1:8	5:8	6:8
Yellow ECOR1-3 x MSE1-4	0:8	0:8	1:8

Figure 2: Three different concentrations were used to sequence the restriction enzyme fragments from the eight pale poppy samples using the LIZ500 size concentration.

Size Standard	LIZ600		
Concentration	12.5%	25%	50%
Primer Set Colors (success rate):	7:8	7:8	6:8
Blue ECOR1-1 x MSE1-5			
Green ECOR1-2 x MSE1-2	7:8	7:8	6:8
Yellow ECOR1-3 x MSE1-4	7:8	3:8	1:8

Figure 3: Three different concentrations were used to sequence the restriction enzyme fragments from the eight pale poppy samples using the LIZ600 size concentration.

Size Standard	LIZ500	LIZ500	LIZ500
Concentration	6.25%	12.5%	25%
Ratio (success rate)	2:24	12:24	13:24
Size Standard	LIZ600	LIZ600	LIZ600
Concentration	12.5%	25%	50%
Ratio (success rate)	21:24	17:24	13:24

Figure 4: The ratios of successful primer pairs were analyzed after the AFLP protocol.

RESULTS (cont.)

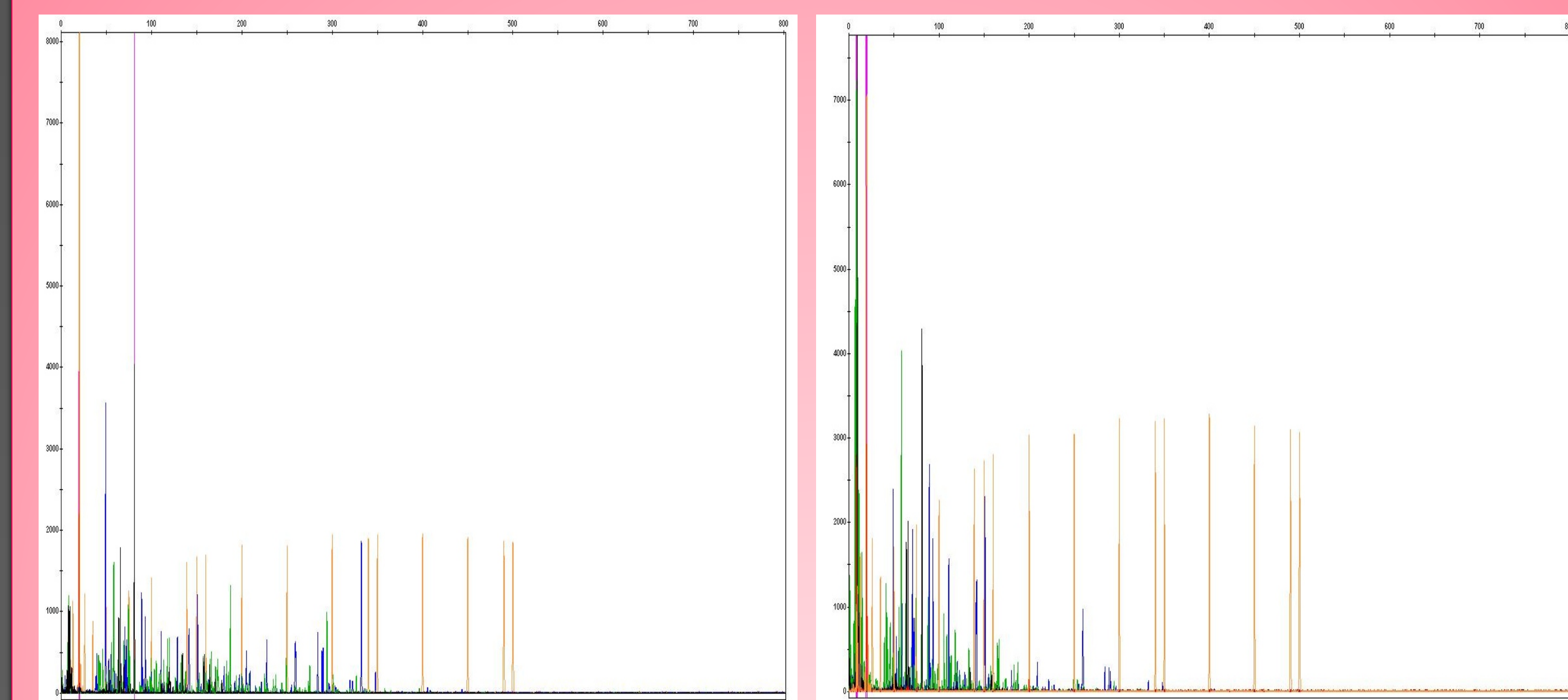


Figure 5: Left: Graph shows a successful range of restriction enzyme fragments for sample 25 at 25% DNA concentration using the LIZ500 size standard. Right: Graph shows a wide range of overshoot to flat line levels of DNA at the LIZ500 size standard for sample 35 at 25% DNA concentration.

DISCUSSION

After testing the AFLP method with two size standards (LIZ500 and LIZ600) and four different DNA concentrations (6.25%, 12.5%, 25%, and 50%), the DNA dilution at 25% in the LIZ500 (primer pairs ECOR1-1 x MSE1-5 and ECOR1-2 x MSE1-2) and at 12.5% in the LIZ600 (all 3 primer pairs) were the most successful.

In the LIZ500 size standard, the yellow primer pair (ECOR1-3 x MSE1-4) lacked any success and was declared as a failed primer.

CITATIONS

Collet, Dominique M. "A Survey of Two Sensitive Plant Taxa *Papaver alboroseum* and *Arinca lessingii* ssp. *Norbergii*, in Portage and Bear Valleys, Chugach National Forest, Alaska." *USDA Forest Service*. (2005): 3-18.

Vos, Pieter, Rene Hogers, Marjo Bleeker, Martin Reijans, Leo van de Lee, Miranda Hornes, Adrie Frijters, Jerina Pot, Johan Peleman, Martin Kuiper and Marc Zabeau. "AFLP: a new technique for DNA fingerprinting." *Oxford University Press*. 23.21 (1995): 4407-13.

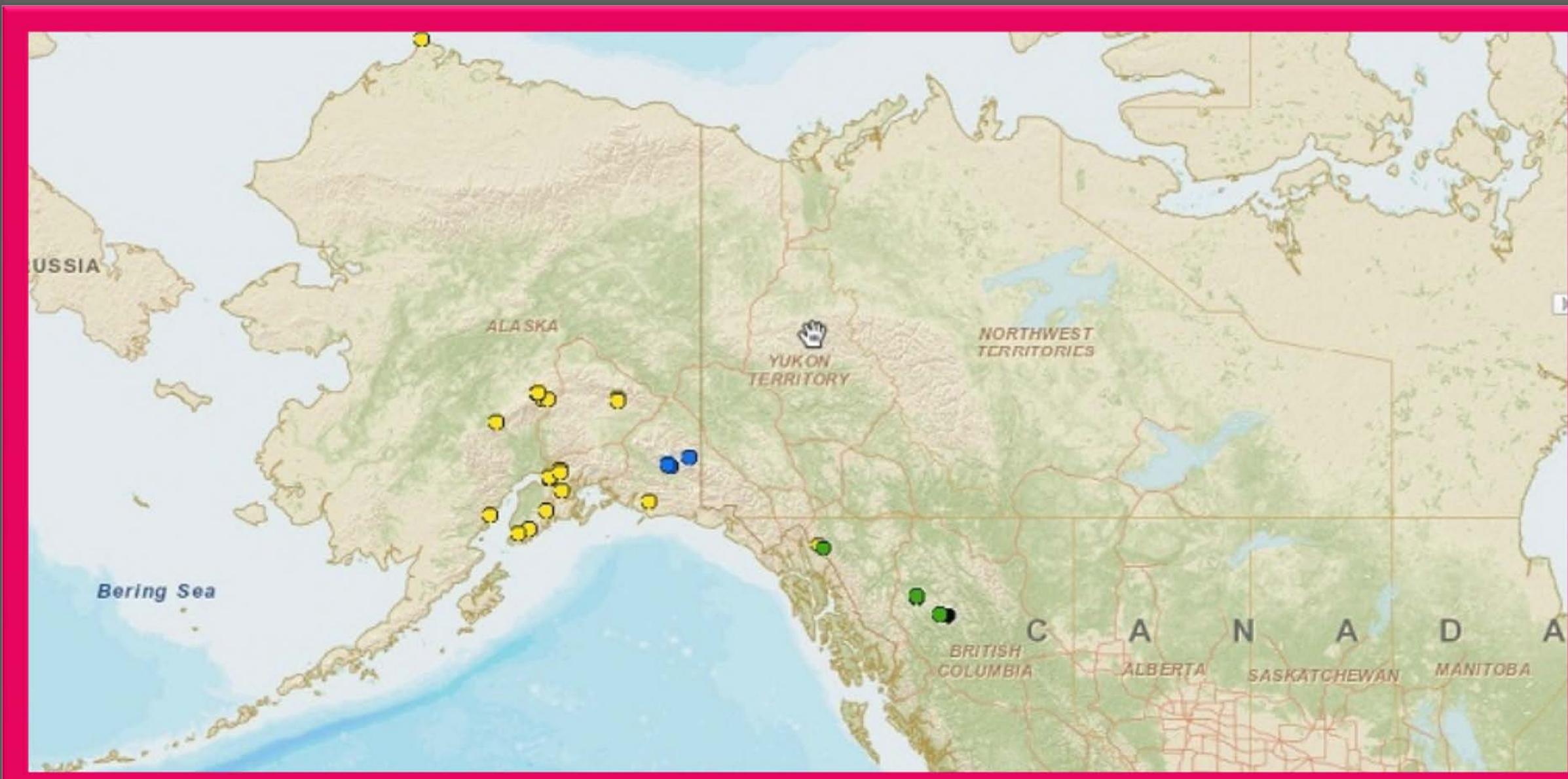


Figure 1: Distribution of *Papaver alboroseum*. Map. *E-Flora BC: Electronic Atlas of the Plants of British Columbia*. 2012.

METHODS

In this project, the Amplified Fragment Length Polymorphism (AFLP) method was used (Vos *et al*, 1995).

Here are the steps:

1. Restriction Enzyme Digest
2. Adaptor preparation & ligation
3. PCR 1- Pre-amplification
4. PCR 2 - Selective amplification
5. Formamide and denature sequencing plate
6. Data analysis

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