

UNDERGRADUATE RESEARCH & SCHOLARLY ACTIVITY

Research Objective

Examine variations in Rainbow smelt otolith core and edge ⁸⁸Sr concentrations across marine and freshwater sites.



Introduction

This study aims to understand the life history of Rainbow Smelt Osmerus mordax in the nearshore waters of the Beaufort Sea, a species that is data limited throughout the state of Alaska. Understanding Rainbow Smelt life history is critical for creating a holistic picture of the Arctic marine ecosystem. The Rainbow Smelt found in the Beaufort Sea and surrounding estuaries are anadromous³, breeding and spawning in freshwater, with adults being found in rivers, estuaries, and marine waters. They are an important cultural resource as they serve as a food source for predators, like Ringed Seals⁴ Pusa hispida and Chinook Salmon² Oncorhynchus tshawytscha. Our objective is to analyze Rainbow Smelt otolith ⁸⁸Sr concentrations and ^{86/87}Sr ratios to gain a better understanding of their life history, using a Laser Ablation Multi Collector Inductively Coupled Mass Spectrometer (LA-MC-ICPMS) system. This beam-based method uses small beam diameters that allows for high spatial resolution and will help produce detailed chronologies of isotopic variation over the fish's lifetime. We hypothesize that otoliths from marine sites will exhibit higher ⁸⁸Sr concentrations and ^{86/87}Sr ratios at the edge of the otolith compared to freshwater sites.

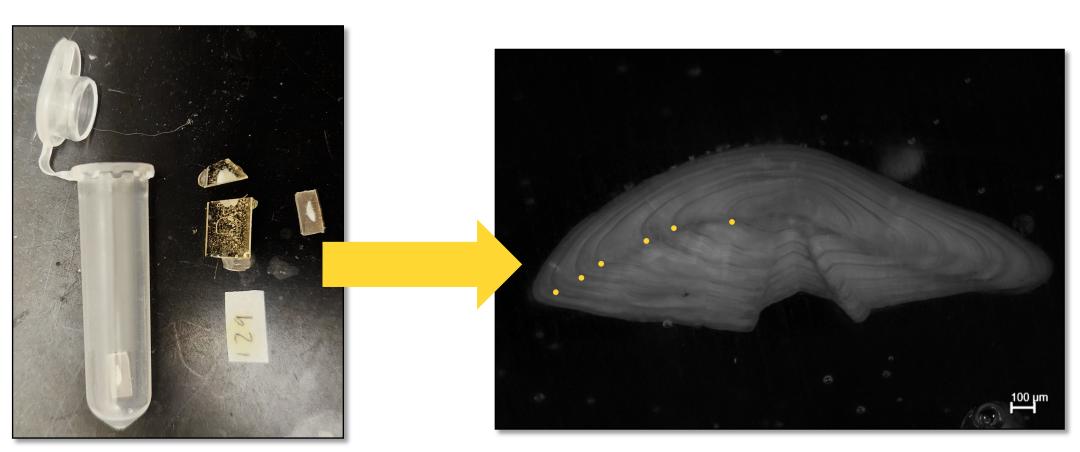


Figure 2.– Thin section of whole otoliths are digitized for aging. This fish was aged at six years old.

Assessing ⁸⁸Sr Concentration and ^{86/87}Sr Ratio Variations in Rainbow Smelt Otoliths using Laser Ablation Mass Spectrometry

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Methods

- Rainbow Smelt samples were collected in the summer months of 2022 (n=150) and 2023 (n=177). Data gathered included: total length, total weight, sex, sexual maturity, and sagittal otolith removal.
- A subsample of otoliths (n=80) will be taken for stable isotope analysis (SIA) to investigate ⁸⁸Sr concentrations. Subsamples are chosen to represent various life histories in both males and females.
- Preparation of samples for SIA is in conjunction with fish aging methods where otoliths are encapsulated in epoxy resin, cut transversely into a thin sections and aged by counting the annuli on the otolith as depicted in Figure 3. An additional step of mounting the thin section for stable isotope analysis then occurs.
- Figure 3 illustrates the process of preparing individual samples and mounting multiple otoliths for SIA to streamline processing. By increasing the number of otoliths that can be loaded into the instrument, the total time and cost of beam-based analyses can be reduced¹.

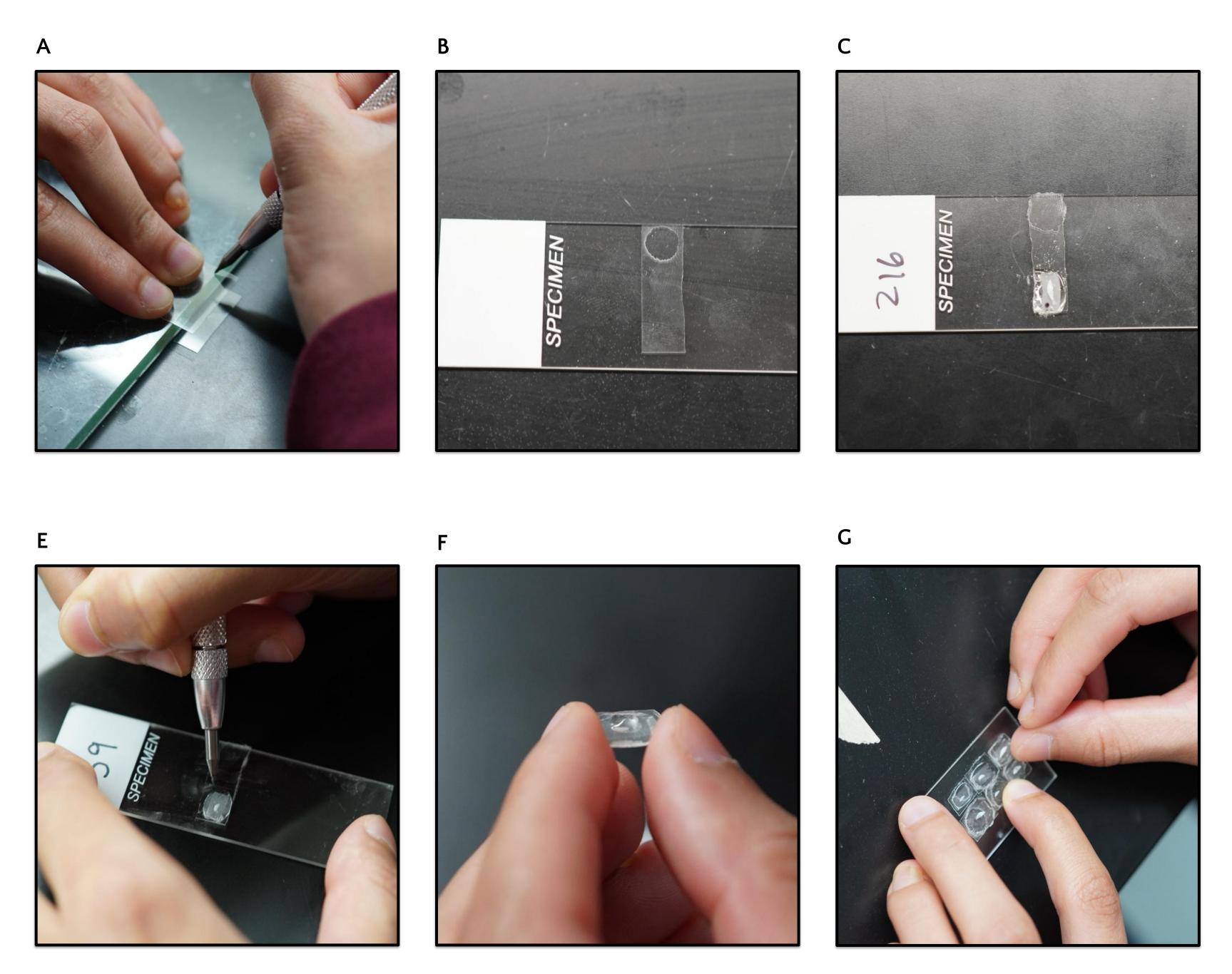
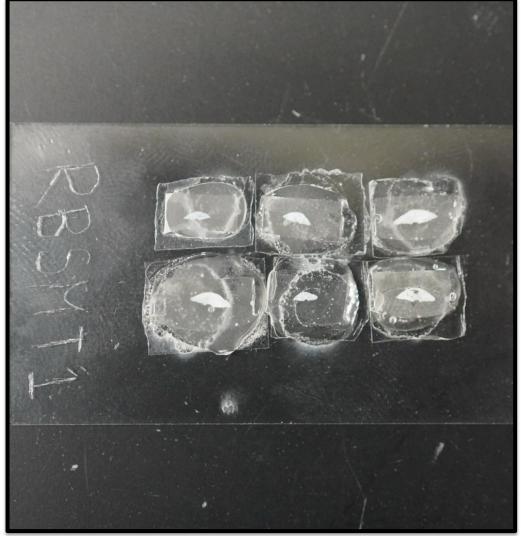


Figure 3.- To mount multiple otoliths for beam-based microchemical analysis, A score a cover glass with a diamond tipped scribe to slice into three strips, **B** attach one end of the glass strip to a microscope slide, **C** mount the otolith thin section in thermoplastic resin on the free edge of the glass slip, **D** polish the mounted otolith using 800 and 3000 grit paper, **E** score the glass strip to cut and free the otolith from the slide, F apply cyanoacrylate glue to free otolith mount, G and affix otolith to slide. Petrographic slides H can hold up to nine otoliths for stable isotope analysis.





We will be using the LA-MC-ICPMS system the week of April 14, 2024, to measure the ⁸⁸Sr concentrations and ^{86/87}Sr ratios of 80 Rainbow Smelt otoliths. We will graph the data from the spectrometry in R and analyze potential life history trends. Continued yearly data collection of Rainbow Smelt otoliths in the future could increase our understanding of their life history, as well as the potential current and future effects of climate change on their habitat usage.

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COLLEGE OF FISHERIES AND OCEAN SCIENCES

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Future Research

Acknowledgements

References

¹ Donohoe, C.J., and C.E. Zimmerman. 2010. A method of mounting multiple otoliths for beam-based microchemical analyses. Environmental Biology of Fishes 89:473–477.

² Honeyfield, D.C., J.M. Murphy, K.G. Howard, W.W. Strasburger, and A.C. Matz. 2016. An exploratory assessment of thiamine status in western Alaska Chinook salmon (Oncorhynchus tshawytscha). North Pacific Anadromous Fish Commission 6:21–31.

³U.S. Geological Survey. 2024. Nonindigenous Aquatic Species Database. (Osmerus mordax) [online database] U.S. Geological Survey, Gainesville, Florida. Available: <u>nas.er.usgs.gov</u>

⁴ U.S. Office of the Federal Register. 2022. Designated critical habitat. Code of Federal Regulations, Title 50, Part 226. U.S. Government Printing Office, Washington, D.C.