



### Grayling and eDNA

- Arctic Grayling (*Thymallus arcticus*) freshwater species
  - Broad distribution across Alaska
- Opportunistic feeders; wide diet variety
- Follow spawning salmonid to feed
- Not an important species for subsistence harvest
- Popular for sport fishing; various colors and sizes
- Environmental DNA analysis
- Potential source of information of distribution, abundance, and ecology
- qPCR (quantitative Polymerase Chain Reaction) strategy for extracted DNA to determine relative amount of species specific DNA
- Will we find an abundance of Arctic Grayling DNA in the Chena **River during the salmon spawning season?**



Image 1. Greg Beutler fly fishes for Arctic Grayling off Chena Hot Springs Road, close to the North Fork sample site Photo:John Fillmore

### Methods

- Collected water samples with Citizen Science Sampler • 7 locations on 3 separate days: 3 samples each sample site
- Isolated DNA from filters
- Prepared DNA for quantification
- Processed species specific assay with PCR
- Processing samples through qPCR machine (qTOWER3 84)

# **Assessing Arctic Grayling Relative Abundance and Distribution Through Environmental DNA Analysis**

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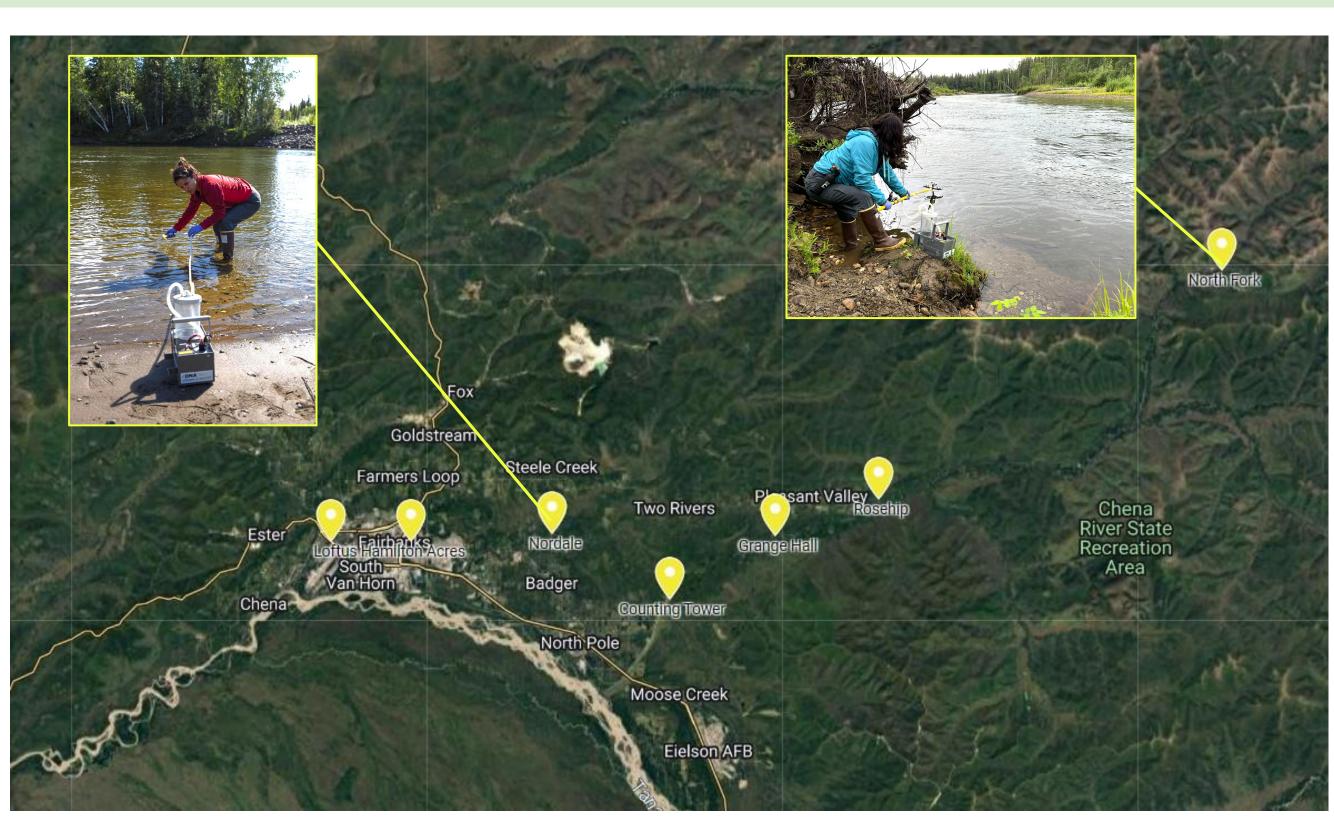


Image 2. Chena River Sample Sites: North Fork, Rosehip, Grange Hall, Moose Creek Counting Tower, Hamilton Acres, Loftus and Nordale. Photos: Maggie Harings, UAF

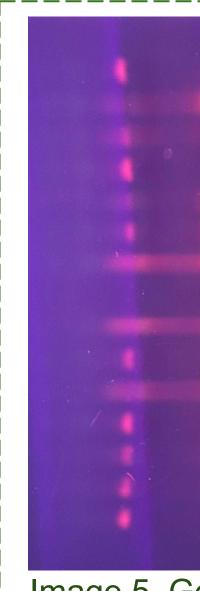


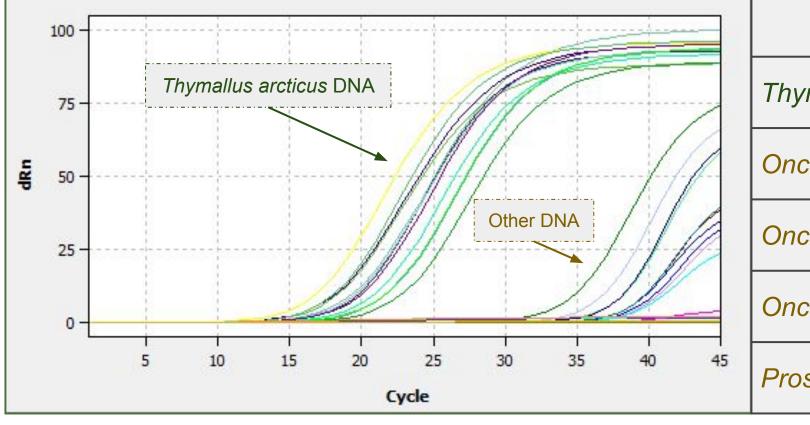
Image 3. Water sample filters before eDNA extraction

### Table 1. DNA from tissue samples

Thymallus arcticus	Arctic Grayling
Coregonus autumnalis	Arctic Cisco
Oncorhynchus keta	Chum Salmon
Prosopium cylindraceum	Round Whitefish
Dallia pectoralis	Alaska Blackfish
Cottus cognatus	Slimy Sculpin
Oncorhynchus tshawytscha	Chinook Salmon
Oncorhynchus mykiss	Rainbow Trout/ Steelhead
Coregonus sardinella	Least Cisco









## We acknowledge the Alaska Native nations located on the traditional lands of the lower Tanana River from which we collected our environmental samples from.

Image 4. Extracting DNA from filters Photo: Alice Bailey, UAF

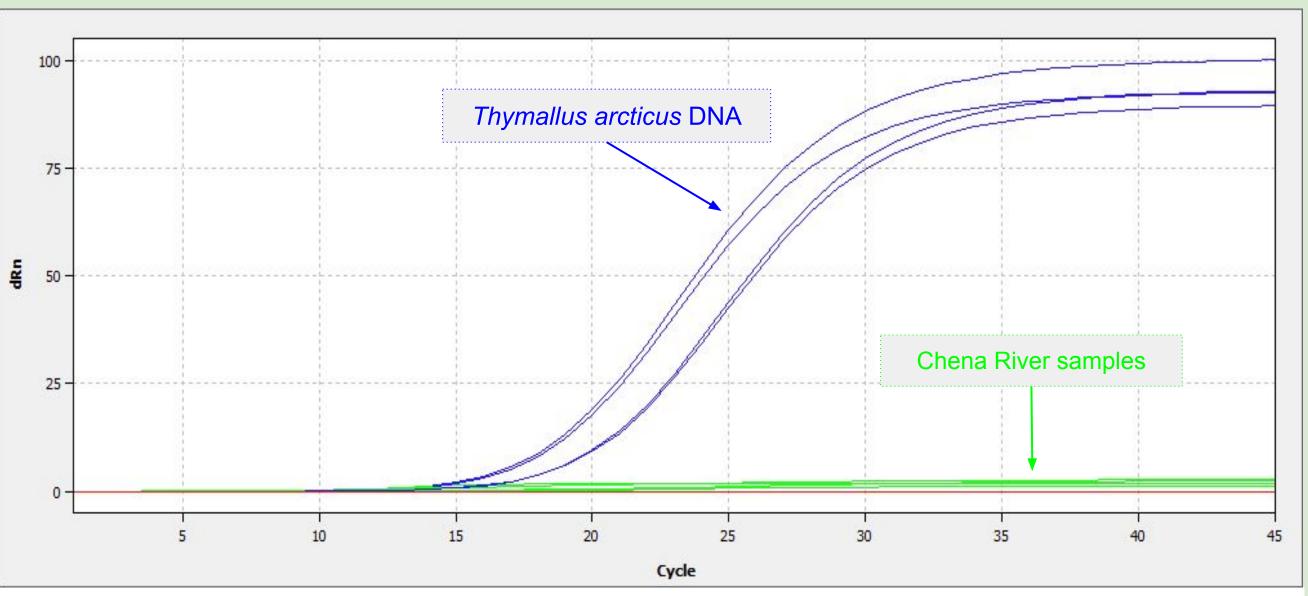
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₋adder Negative Thymallus arcticus Coregonus autumnalis Oncorhynchus keta Prosopium cylindraceum Dallia pectoralis Thymallus arcticus Ladder Thymallus arcticus Cottus cognatus Thymallus arcticus Oncorhynchus tshawytscha Oncorhynchus mykiss Coregonus sardinella Negative Ladder

### Image 5. Gel electrophoresis from PCR

Species	Ct Values	
ymallus arcticus	14.97 - 17.31	
ncorhynchus keta	38.1	
ncorhynchus mykiss	38.65	
ncorhynchus tshawytscha	36.52	
osopium cylindraceum	32.95	

- low to amplify (Figure 2).



- too low to amplify.
- Next Steps:

  - spawning periods

# 10.1007/s12686-017-0883-1

Schoen, E. R., Sellmer, K. W., Wipfli, M. S., López, J. A., Ivanoff, R., & Meyer, B. E. (2022). Piscine predation on juvenile salmon in sub-arctic Alaskan rivers: Associations with season, habitat, predator size and streamflow. *Ecology of* Freshwater Fish, 31, 243–259. https://doi.org/10.1111/eff.12626

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### **Preliminary Analysis**

 Tested assay specificity with tissue samples from Arctic Grayling and closely related/unrelated species (Table 1).

 High DNA concentrations of most salmonids indicate amplification during PCR (Image 5) and qPCR (Figure 1 & Table 2). • qPCR indicates DNA concentrations of Chena River samples too

Figure 2. qPCR results of Arctic Grayling vs.Chena River eDNA samples.

### Discussion

• Latest qPCR analysis indicates eDNA sample concentrations are

• Further testing assay (Rodgers, et al) with known DNA concentrations to develop level of detection for Arctic Grayling. • Data configured in relation to Chum and Chinook salmon eDNA

• qPCR method may become a complementary technique in the field of assessing relative quantities of species over time and space.

### Sources

Rodgers, T.W., Olson, J.R., Klobucar, S.L. et al. Conservation Genet Resour (2018) 10: 859. https://doi.org/

### Acknowledgements